

SYNTHESIS OF A PROTEIN BIOSYNTHESIS INHIBITOR, 5'-TRI-
PHOSPHORYLADENYLYL-(2'-5')-ADENYLYL-(2'-5')-ADENOSINE

Morio Ikehara, Kazuyuki Oshie and Eiko Ohtsuka

Faculty of Pharmaceutical Sciences, Osaka University

Suita, Osaka, Japan 565

A protein biosynthesis inhibitor, 5'-triphosphoryladenyl-yl-(2'-5')-adenyl-yl-(2'-5')-adenosine was synthesized by polymerization of N⁶-benzoyl-3'-O-(o-nitrobenzyl)adenosine 5'-phosphate followed by reaction with pyrophosphate using 1,1'-carbonyldiimidazole.

Recently an inhibitor of protein biosynthesis in interferon-treated cells has been found and characterized as 5'-triphosphoryladenyl-yl-(2'-5')-adenyl-yl-(2'-5')-adenosine (pppA^{2'}p^{5'}A^{2'}p^{5'}A¹) (I). In this paper we wish to communicate a method of chemical synthesis of the compound I.

N⁶-Benzoyl-3'-O-(o-nitrobenzyl)adenosine (II) [m.p. 218-221°, NMR: δ 4.22 (br. s, 2H, 3'- and 4'-OH), 5.08 (d, 2H, Ar-CH₂)] was obtained by crystallization from the mixture of 2'- and 3'-o-nitrobenzyl derivatives, which were synthesized by benzylation of N⁶-benzoyl-5'-monomethoxytrityladenosine²⁾ with o-nitrophenyldiazomethane.³⁾ The compound II was phosphorylated with POCl₃ according to a standard procedure⁴⁾ and 5'-monophosphate (III) (paper electrophoresis, RpA 0.98) was obtained by chromatography on a DEAE-Sephadex A-25 column in a yield of 64%.

Compound III was then polymerized using dicyclohexyl carbodiimide as condensing reagent. After 15 days at room temperature, the reaction mixture was worked up as usual and applied to a column of DEAE-cellulose (bicarbonate form) and eluted with a linear gradient of triethylammonium bicarbonate buffer (0-0.5M, total 10 l.). The trimer (IV) was obtained in a yield of 7.7% together

with dimer (9.8%), tetramer (6.3%) and higher oligomers (13.2%). For analysis a small amount of samples were irradiated with UV (λ over 280 nm, for 1hr) to remove *o*-nitrobenzyl groups. Each product is characterized separately by comparison of R_f in paper chromatography before and after bacterial alkaline phosphatase treatment⁵⁾, UV absorption properties, hypochromicity and ϵ values as tabulated in Table I.

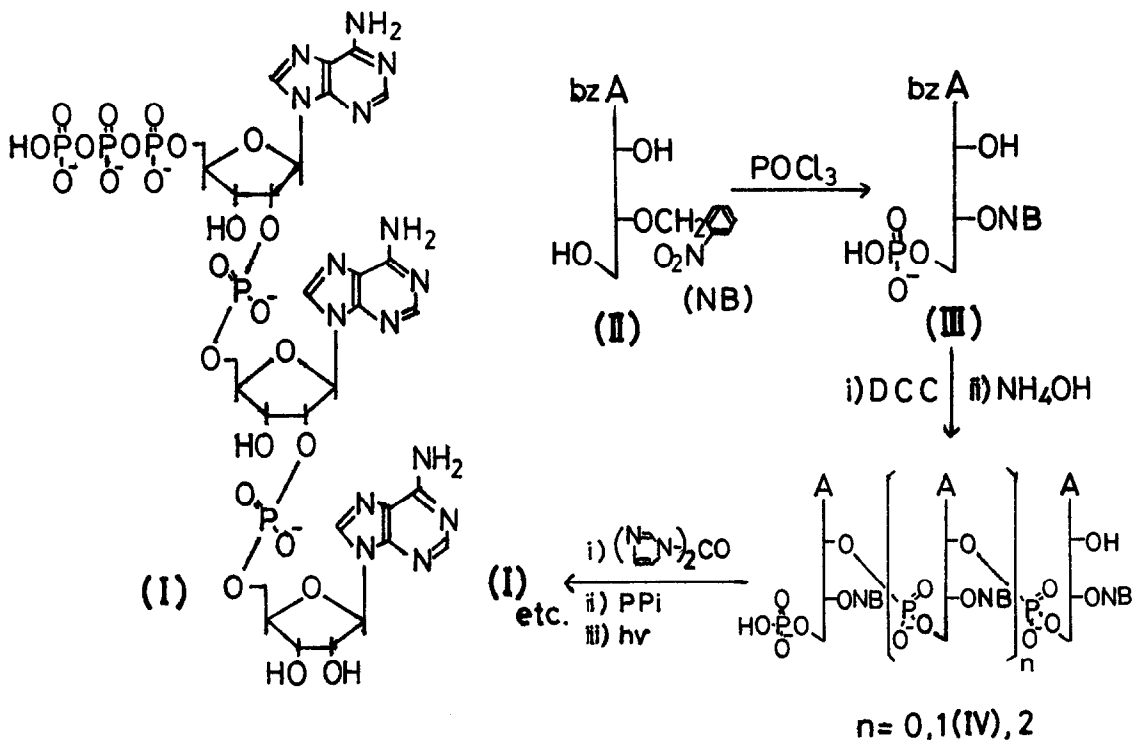


Table I Properties of $\text{pA}(2' \text{ p } 5' \text{ A})_n$

	R_f^a	R_{pA}^a	$R_{\text{A-pA}}^b$	A/pA^c	$\text{H}(\%)^d$	ϵ^e
$\text{pA}(2' \text{ p } 5' \text{ A})$	0.34	0.79	1.03			
$\text{A}(2' \text{ p } 5' \text{ A})$	0.52	1.20		1/1.10	16.1	12900
$\text{pA}(2' \text{ p } 5' \text{ A})_2$	0.21	0.50	1.08			
$\text{A}(2' \text{ p } 5' \text{ A})_2$	0.41	0.95		1/2.08	19.5	12400
$\text{pA}(2' \text{ p } 5' \text{ A})_3$	0.14	0.33	1.13			
$\text{A}(2' \text{ p } 5' \text{ A})_3$	0.28	0.65		1/3.13		

(Table I continued)

a. Solvent : n-propanol-conc. ammonia-water (55 : 10 : 35); b. Relative mobility in paper electrophoresis at pH 7.5 as A assumed to be R_m 0.0 and pA as 1.0; c. Ratio of A and pA after venom phosphodiester treatment and paper electrophoresis; d. hypochromicity estimated by alkaline hydrolysis; e. ξ value calculated from ξ of pA assumed to be 15400.

Compound IV (triethylammonium salt) was then converted to pyridinium salt by passing through a Dowex 1x2 column (pyridinium form) and dried thoroughly by coevaporation with anhydrous pyridine several times. The residual IV was dissolved in DMF and allowed to react with 1,1'-carbonyldiimidazole⁶) (10 equiv.) at room temperature for 14 hrs. After decomposition of unreacted reagent with MeOH, a solution of tri-n-butylammonium pyrophosphate (10 equiv.) in DMF was added and the mixture was kept at room temperature for 24 hrs. After removal of precipitate (probably imidazolium phosphate) by filtration, DMF was evaporated off in vacuo. The residue was dissolved in water and irradiated with UV light over wavelength of 280 nm for 3 hrs. Compound I was isolated by successive paper chromatography and paper electrophoresis in a yield of 40%. The di- and tetramer were obtained by the essentially same method. Properties of these products are summarized in Table II.

Table II Properties of pppA(2' p 5' A)_n

	Rf ^a (R _{pA})	Rf ^b (R _{pA})	Rf ^c (R _{pA}) (R _{Ap})	R _{A-pA} ^d	ξ	H(%) ^e
pppA(2' p 5' A)	0.18 (0.58)	0.22 (1.05)	0.16(0.52) (0.46)	1.25	12900	16.1
pppA(2' p 5' A) ₂	0.11 (0.35)	0.09 (0.43)	0.10(0.33) (0.29)	1.34	11500	25.3
pppA(2' p 5' A) ₃	0.07 (0.23)	0.03 (0.14)	0.06(0.18) (0.17)	1.40	104000	32.4

a. Solvent, n-propanol-conc. ammonia-water (55 : 10 : 35); b. sat. (NH₄)₂SO₄ 600g in 0.1M sodium phosphate buffer (pH 6.8) + n-propanol (20 ml); c. n-propanol-conc. ammonia-water (60 : 10 : 30); d. R_{A-pA} stands for relative mobility in paper electrophoresis (pH 7.5) assumed A as R_f 0.0 and pA as 1.0; e. hypochromicity as calculated from alkaline hydrolysis.

Thus the compound I, $\text{pppA}^{2'}\text{p}^{5'}\text{A}^{2'}\text{p}^{5'}\text{A}$ and related compounds were proved to be identical to natural protein inhibitors obtained previously.¹⁾

References

1. I. M. Kerr and R. E. Brown, Proc. Nat. Acad. Sci. U. S. A., 75, 256 (1978).
2. R. H. Hall, Biochemistry, 3, 769 (1964).
3. E. Ohtsuka, T. Tanaka, T. Wakabayashi, Y. Taniyama and M. Ikehara, Abstracts of papers presented at the 5th Symposium on Progress in Reaction and Synthesis, p. 93 (1978).
4. M. Yoshikawa, T. Kato and T. Takenishi, Bull. Chem. soc. Jap., 42, 1022 (1969).
5. E. Ohtsuka, H. Tsuji and M. Ikehara, Chem. Pharm. Bull., 22, 1022 (1974).
6. D. E. Hoard and D. G. Ott, J. Am. Chem. Soc., 87, 1785 (1965).

(Received in Japan 16 June 1979)